Changes in antimicrobial resistance, serotypes and genotypes in Streptococcus pneumoniae over a 30-year period

J. Liñares^{1,2}, C. Ardanuy^{1,2}, R. Pallares^{2,3} and A. Fenoll⁴

1) Microbiology, Hospital Universitari de Bellvitge and Universitat de Barcelona-IDIBELL, L'Hospitalet de Llobregat, Barcelona, 2) CIBER de Enfermedades Respiratorias, Instituto de Salud Carlos III, Madrid, 3) Infectious Diseases Departments, Hospital Universitari de Bellvitge and Universitat de Barcelona-IDI-BELL, L'Hospitalet de Llobregat, Barcelona and 4) Centro Nacional de Microbiología, Instituto de Salud Carlos III, Majadahonda, Madrid, Spain

Abstract

Over the past three decades, antimicrobial resistance in Streptococcus pneumoniae has dramatically increased worldwide. Non-susceptibility to penicillin in S. pneumoniae was first described in Australia in 1967, and later in New Guinea (1974), South Africa (1977), and Spain (1979). Most of these strains showed resistance to multiple antibiotics and belonged to serotypes 6A, 6B, 19A, 19F, and 23F. By the late 1980s and 1990s, the emergence and rapid dissemination of antibiotic-resistant pneumococci was observed in southern and eastern Europe, North America, South America, Africa, and Asia. Great geographical variability, both in serotype distribution and in the prevalence of resistant pneumococci, has been reported. However, the highest rates of resistance to penicillin and erythromycin worldwide were found in serotypes 6B, 6A, 9V, 14, 15A, 19F, 19A, and 23F. The introduction of the seven-valent pneumococcal conjugate vaccine (PCV7) in the 2000s and a reduction in antimicrobial use were associated with a significant decline in the incidence of invasive pneumococcal infections and in rates of antibiotic resistance in the USA. However, an increase in the incidence of infections caused by non-PCV7 serotypes, especially multiresistant serotype 19A pneumococci, has been observed in many countries over the last 5 years. The dynamic character of serotypes and antibiotic resistance in S. pneumoniae should be controlled by a policy of prudent antibiotic use and by implementation of the new generation of conjugate vaccines.

Keywords: Genotypes, penicillin-resistant, review, serotypes, Streptococcus pneumoniae Clin Microbiol Infect 2010; 16: 402–410

Corresponding author and reprint requests: J. Liñares, Servicio de Microbiología, Hospital universitario de Bellvitge, Feixa Llarga s/n, 08907 L'Hospitalet de Llobregat, Barcelona, Spain E-mail: fina.linares@bellvitgehospital.cat

Introduction

Streptococcus pneumoniae is the most important pathogen in otitis, sinusitis, bronchitis, and community-acquired pneumonia, as well as a predominant cause of meningitis and bacteraemia. Before 1967, this pathogen was uniformly susceptible to penicillin and most other antimicrobial agents. The first penicillin-non-susceptible pneumococcus was described in Australia in 1967; it had an MIC of penicillin of 0.6 mg/L, and it also showed resistance to tetracycline (MIC 5 mg/L) [1]. In 1974, 12% of 518 New Guinean isolates were penicillinnon-susceptible pneumococci (PNSP) [2]. Pneumococci with much higher resistance to penicillin and other antibiotics were first detected in 1977 in South Africa [3,4]. These multidrug-resistant pneumococci of serotypes 6A and 19A

had penicillin MICs ranging from 0.12 to 4 mg/L, and were isolated from hospitalized paediatric carriers (29% of 543), causing bacteraemia in four patients [4]. In Spain, the first PNSP strain with a penicillin MIC of 0.5 mg/L was isolated from the blood from an adult patient in a hospital in Barcelona in 1979, and a year later one invasive isolate with a penicillin MIC of 2 mg/L was detected in the same hospital in Barcelona [5]. By the 1980s, a high prevalence of antibiotic resistance among pneumococci was being reported in other countries, leading to serious therapeutic problems, mainly in the treatment of pneumococcal meningitis [3,4,6–10]. The emergence and rapid dissemination of antibiotic-resistant pneumococcal clones in areas of southern and eastern Europe, North America, South America and Asia in the 1990s was associated with an increase in antibiotic consumption [11,12].

Although 92 pneumococcal serotypes have been described, the highest penicillin and erythromycin resistance proportions worldwide were associated with serotypes 6B, 6A, 9V, 14, 15A, 19F, 19A, and 23F, the so-called 'paediatric serotypes'. The introduction of the seven-valent pneumococcal conjugate vaccine (PCV7) for children in the 2000s, which included serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F, was associated with a significant decline in penicillin resistance rates in S. pneumoniae in many countries [13–16]. However, an increase in the incidence of non-PCV7 serotypes, especially multiresistant serotype 19A strains, has been observed in many countries over the last 5 years [16–22].

This review analyses geographical trends in antibiotic resistance and the serotypes/genotypes in S. pneumoniae over the last three decades. It also includes the evolution of penicillin, erythromycin and fluoroquinolone resistance among invasive pneumococci received at the Spanish Reference Laboratory for Pneumococci over a 30-year period (1979– 2008).

Resistance to β -Lactam Antibiotics

The mechanism of penicillin resistance in S. pneumoniae involves structural changes in the penicillin targets, the penicillin-binding proteins 1A, 2X, and 2B [23]. These changes result in reduced affinity for penicillin as well as for other β lactam antibiotics. However, ceftriaxone, cefotaxime and carbapenems are less affected, and are generally the most active compounds [24].

Previous studies have demonstrated that PNSP (MICs \geq 0.12 mg/L) can be associated with treatment failures in patients with meningitis, because the cerebrospinal fluid levels achieved with penicillin or the standard dosage of third-generation cephalosporins are insufficient to eradicate the infecting organism. The classical NCCLS breakpoints for penicillin (susceptible, ≤0.06 mg/L; intermediate, 0.12–1 mg/L; and resistant, ≥ 2 mg/L) were established in the late 1970s in order to prevent failures in patients with pneumococcal meningitis caused by PNSP. In contrast, S. pneumoniae strains with penicillin MICs of 0.12–2 mg/L had little effect on outcome in patients with pneumonia and other non-meningeal systemic pneumococcal infections who were treated with parenteral penicillin, amoxycillin, cefotaxime, or ceftriaxone [25-31]. According to these observations, the CLSI breakpoints for ceftriaxone and cefotaxime were modified in 2002, distinguishing between meningeal infections (susceptible, MIC ≤0.5 mg/L; intermediate, MIC 1 mg/L; and resistant, $MIC \geq 2$ mg/L) and non-meningeal infections (susceptible, $MIC \leq 1$ mg/L; intermediate, MIC 2 mg/L; and resistant,

MIC \geq 4 mg/L). However, penicillin breakpoints were not modified until 2008, when the site of infection and route of administration were considered. The current penicillin parenteral breakpoints for non-meningeal infections are $MIC \leq 2$ mg/mL (susceptible), MIC 4 mg/mL (intermediate), and MIC \geq 8 mg/mL (resistant), whereas MICs \geq 0.12 mg/L for strains causing meningeal infection are considered to reflect resistance [32].

In the USA, S. pneumoniae was uniformly susceptible to penicillin until 1987. After that, rates of PNSP with a penicillin MIC \geq 0.1 mg/L progressively increased from very low rates (5%) before 1991 to 14% in 1993–1994 [33,34]. In 1997, among 3237 invasive isolates, 25% were PNSP, and among these, 13.6% had a penicillin MIC \geq 2 mg/L [35].

Global surveillance studies have shown that β -lactam-nonsusceptibility rates increased worldwide during the 1990s and 2000s. The Alexander Project monitored resistance in S. pneumoniae from 1992 to 2001, and reported increases in the levels of non-susceptibility to penicillin from 24.9% in 1992 to 30.2% in 2001 in Spain, from 7.7% to 35.8% in France, and from 5.6% to 20.4% in the USA, whereas in Italy, Germany, and the UK, resistance rates remained below 5% during this period [36].

The PROTEKT US study, with 39 495 pneumococcal isolates from patients with community-acquired respiratory tract infections in the USA, showed that intermediate non-susceptibility to penicillin (MIC 0.12–1 mg/L) increased from 12.5% in 2000–2001 to 20.0% in 2003–2004, whereas penicillin resistance (MIC \geq 2 mg/L) declined from 26.3% in 2000– 2001 to 16.5% in 2003–2004. However, the percentage of resistance to amoxycillin (MIC \geq 8 mg/L) remained low and stable, from 4.4% in 2000–2001 to 4.1% in 2003–2004 [37]. The PRO-TEKT study also analysed the antibiotic susceptibility of 20 142 pneumococcal isolates collected worldwide from respiratory tract infections in 2001–2004. The highest penicillin-non-susceptiblity rates were found in South Africa (74%), the Far East (63%), and the Middle East (54%). In southern European countries, rates of PNSP were higher than those found in northern European countries. The highest rates of PNSP (intermediate and resistant isolates) were found in France (40.4% and 15.9%), Greece (42.0% and 15.9%), and Spain (29.4% and 13.1%) [38]. In seven Latin American countries, a study of 1561 pneumococcal isolates collected from 1997 to 2001 revealed a global rate of PNSP of 30.7%, ranging from 25% in Mexico to 2.8% in Venezuela. Resistance to penicillin (MIC \geq 2 mg/L) and cefotaxime (MIC \geq 4 mg/L) was found in 11.9% and 0.4% of isolates, respectively [39]. Southern and eastern countries of the Mediterranean region reported that 26% (335) of the 1298 invasive isolates studied were PNSP, with the highest proportions being reported in Algeria (44%) and Lebanon (40%) [40].

The European Antimicrobial Resistance Surveillance System (EARSS), an international network of national surveillance systems that has been collecting antimicrobial susceptibility testing data on invasive S. pneumoniae isolates since 1999, also reported that most northern European countries had levels of PNSP below 5%, whereas southern and eastern European countries had PNSP levels above 25% [41]. In 2008, 1152 (10%) of the 11 584 invasive S. pneumoniae isolates reported to the EARSS by 32 countries were PNSP [41]. Among the northern European countries, only in Finland and Ireland had the frequency of PNSP risen significantly from 2005 to 2008 (7% (37/525) vs. 11% (71/642), and 11% (43/397) vs. 23% (101/441), respectively). In contrast, four countries with the highest levels of PNSP in the early 2000s (France, Spain, Belgium, and Israel) showed significantly decreasing rates of PNSP over the subsequent years.

Data from the Spanish Reference Laboratory for Pneumococci are shown in Fig. 1. A total of 25 166 invasive pneumococci were received during the period 1979–2008. Rates of invasive PNSP isolates progressively increased from 6% in 1979 to 44.4% in 1989 (p <0.001), and the proportion of resistant isolates (MIC \geq 2 mg/L) increased from 0% to 15.4% during this period (p <0.001). During the period 1990–1998, the proportion of PNSP plateaued, oscillating between 34.5% and 43.7%. In the last decade, rates of PNSP progressively decreased from 33.9% in 1999 to 22.3% in 2008 (p <0.001). This decline was especially marked in 2005–2008, and was associated with the implementation of PCV7 for children (Table 1). Moreover, among 1397 pneumococcal isolates from cerebrospinal fluid, non-susceptibility rates for cefotaxime (MIC \geq 1 mg/L) decreased from 21.7% in 2000 to 10.5% in 2008 [16].

In a study performed in 33 US medical centres, the rate of ceftriaxone non-susceptibility (MIC \geq 2 mg/L) decreased from 14.4% among 1531 pneumococcal isolates collected in 1999–2000 to 5.9% among 1647 pneumococcal isolates collected in 2004–2005 in 41 medical centres [42,43].

In spite of these alarming penicillin resistance levels, pneumococci with penicillin or cefotaxime/ceftriaxone MICs \geq 4 mg/L are rarely described worldwide. If the revised CLSI breakpoints for parenteral penicillin are applied to pneumococci isolated from non-meningeal infections, more than 95% of invasive pneumococcal isolates collected wordwide are currently susceptible to penicillin and third-generation cephalosporins [32]. These antibiotics should continue to be first-line therapy for these infections [25–27,31].

Macrolides

The most frequently encountered mechanism of macrolide resistance in pneumococci is target site modification mediated by the ermB-encoded methylase, conferring resistance to all macrolides, lincosamides, and streptogramin B (MLS_B phenotype). The expression of this phenotype can be constitutive or inducible, becoming active only in the presence of inducing macrolides. The ermB gene is the major cause of macrolide resistance in most European countries,

FIG. I. Trends in penicillin and erythromycin resistance among 19 266 invasive pneumococcal isolates in patients of all ages in Spain (1979– 2008). Ery-R, erythromycin resistance; PCV7, seven-valent pneumococcal conjugate vaccine.

TABLE 1. Trends in antibiotic resistance of 19 266 invasive pneumococcal isolates (all ages) tested at the Spanish Reference Laboratory for Pneumococci according to the old and the new breakpoints for penicillin of the CLSI

IPD, invasive pneumococcal disease. ^aChildren: <14 years old.

b p <0.001 when comparing 1997–2004 vs. 2005–2008.

especially Belgium, France, Poland, Italy, and Spain [38]. The second most common mechanism is mediated by an efflux pump codified by the mef genes (mefA, mefE). This mechanism confers the M phenotype, with resistance to 14-membered and 15-membered ring macrolides, but not to lincosamides or streptogramins. The M phenotype isolates predominate in the USA, Canada, the UK, Germany, and Norway. Less common macrolide resistance could be due to mutations in the 23S rRNA and/or alterations in ribosomal proteins (L4 and L22).

The prevalence of macrolide resistance mechanisms differs considerably among countries, as shown in Table 2 [44–58]. The emergence of pneumococci that carry both ermB and mefE macrolide resistance genes is a cause for concern, especially in Asian countries, Russia, South Africa, and the USA [52,59]. Both genes (ermB and mefE), as well as the tetracycline resistance determinant (tetM), have been related to the composite element Tn2010, which is present in most multidrug-resistant isolates of serotype 19A of clonal complex 320 [18,60].

Macrolide-resistant pneumococci were first detected in 1967 in Canada, but rates of macrolide resistance among pneumococci remained low worldwide (<5%) during the 1970s [3]. By the early 1980s, the highest prevalence of erythromycin-resistant pneumococci was found among pneumococci isolated from hospital carriers in South Africa (63%), whereas, among invasive isolates, the prevalence of resistance was 8.3% in 1983. The majority of these strains showed multidrug resistance [3]. In France, macrolide resistance rates dramatically increased from 0% before 1976 to a

peak of 26% in 1985 among clinical isolates at two hospitals in Paris [61]. In Spain, resistance rates increased from 0% in 1979–1980 to 9.4% in 1990 in a Barcelona hospital [9]. Thereafter, a rapid worldwide increase in the prevalence of macrolide resistance associated with an increase in macrolide consumption, especially of long-acting macrolides such as clarithromycin and azithromycin, was observed [38,62–64].

Global surveillance studies have shown that macrolide resistance rates increased during the 1990s. The Alexander Project gave a global rate of macrolide resistance of 16.5– 21.9% in 1996–1997, increasing up to 24.6% in 1998–2000 [36]. The PROTEKT study showed an overall rate of 31.0% in 1999–2000, increasing to 37.2% in 2003–2004, but there was marked geographical variability [37]. The highest rates (80%) were recorded among isolates collected in the Far East, followed by South Africa (54%) and southern Europe (37%), whereas resistance was lowest in Latin America (15%), Australia (18%), and northern Europe (18%). Macrolide resistance in Europe was notably high in isolates collected from Belgium (31.5%), Spain (33.5%), Hungary (39.4%), Italy (40.8%), Greece (51.4%), and France (55.6%).

The EARSS report for 2008 shows high variability in the proportion of macrolide resistance in Europe. Whereas northern European countries, the Czech Republic and Bulgaria reported resistance rates below 5%, Italy, Turkey, France, Hungary and Cyprus reported resistance rates above 25% [41].

In Spain, overall macrolide resistance rates among invasive pneumococci received at the Spanish Reference Laboratory for Pneumococci remained below 5% until 1986, but thereafter resistance genes (%)

Macrolide

increased to 28% in 2001 and decreased to 21.8% in 2008, as shown in Fig. 1 [16]. After the introduction of PCV7 for children in June 2001 in Spain, a significant decline in macrolide resistance among invasive paediatric isolates (42.9% in 2003 vs. 20.8% in 2006, p <0.001) was observed (Table 1), in agreement with previous reports [13,64]. In contrast, erythromycin resistance rates among invasive pneumococci isolated from adult patients remained stable from 1997 to 2008, fluctuating from 21% to 25% (Table 1) [16].

Several studies have associated macrolide resistance with therapy failure [65]. This argues against the empirical use of macrolides for treatment of pneumococcal pneumonia in countries with high rates of macrolide-resistant pneumococci.

Dual Non-susceptibility to Penicillin and Erythromycin

Over the 1990s, an increase in the proportion of pneumococcal isolates with combined non-susceptibility to penicillin and erythromycin was observed. This increase was related to the spread of classic penicillin-resistant clones that have acquired determinants of macrolide resistance, mostly carried by transposons of the Tn916 family [51,66]. Data from the Alexander Project revealed a dramatic increase in the prevalence of combined resistance from 1.8% in 1992 to 32.7% in 2001 in France, from 3.7% to 17.0% in Spain, and from 3.2% to 15.3% in the USA [36].

The last EARSS report noted that the overall rate of dual non-susceptibility remained below 5% in 2008 in Europe. Although the northern European countries reported the lowest dual non-susceptibility levels, a decrease in the level of dual non-susceptibility from 2005 to 2008 was observed in Belgium (9% vs. 6%) and France (32% vs. 25%). Of concern is the increase observed over the last 4 years in the level of dual non-susceptibility in Ireland (from 3% to 12%), Hungary (from 13% to 21%), and Turkey (from 10% to 23%) [41].

As Table 1 shows, in Spain the rate of combined penicillin and erythromycin resistance among paediatric isolates fluctuated between 24% and 35% in 1997–2003, and thereafter showed a significant and progressive decline until it reached 11.5% in 2008 (p <0.05). Among adult isolates, the rate of combined resistance also declined progressively, from 18.9% in 1997 to 11.5% in 2008 (p <0.05) [16].

Quinolones

The main mechanism of resistance to fluoroquinolones in S. pneumoniae is point mutation producing amino acid changes in the quinolone resistance-determining regions of the subunits of DNA topoisomerase IV $(ParC₂ParE₂)$ and DNA gyrase (GyrA₂GyrB₂). However, resistance can also be acquired by intraspecific recombination or by interspecific recombination with streptococci of the mitis group [67–69].

The new, or respiratory, fluoroquinolones such as levofloxacin and moxifloxacin have enhanced activity against pneumococci when compared with the older ones (ciprofloxacin), and have become therapeutic alternatives for the treatment of community-acquired pneumonia in adults, because their spectrum of activity includes S. pneumoniae, Legionella pneumophila, and other atypical pathogens. Increased use of these antimicrobials has been associated with the emergence of resistance in S. pneumoniae in both Canada and Spain [70,71].

A multicentre study performed in Europe in 2004–2005 involving community-acquired respiratory tract infections [72] showed a low level of quinolone resistance in the majority of European countries, with the exception of Poland (4.4%), Finland (6.6%), and Italy (7.2%). Higher rates were also detected in some Asian countries [73], as well as recently in Canada (7.3% in 2006) [74]. In this country, the increase in ciprofloxacin resistance observed between 1998 (0.6%) and 2006 (7.3%) was strongly associated with an increase in fluoroquinolone consumption [74]. In Spain, two nationwide surveillance studies performed in 2002 and 2006 showed a stable rate of ciprofloxacin resistance (2.6% and 2.3%, respectively) [68,69].

There are several reports of treatment failure with the use of quinolones in the treatment of pneumococcal infection caused by fluoroquinolone-non-susceptible isolates. The risk factors identified in these reports were previous fluoroquinolone use, chronic obstructive pulmonary disease, hospitalization, and living in nursing homes [73]. However, once patients with these risk factors were excluded, the efficacy of levofloxacin in the treatment of pneumococcal pneumonia was proven [75]. To avoid therapeutic failures, it is very important to detect strains with first-step mutations, which usually have low-level ciprofloxacin resistance, with MICs of 4–8 mg/L, and frequently appear to be levofloxacin-susceptible, with MICs of 1–2 mg/L [68,69]. These strains could become highly resistant under selective fluoroquinolone pressure. In our experience, the ciprofloxacin breakpoint of 4 μ g/mL, suggested by Chen et al., is the best marker for detecting strains with first-step mutations [68– 70]. These strains, wrongly identified as susceptible, accounted for 2.1% of 665 pneumococcal isolates collected in 2003 during the Canadian Respiratory Organism Susceptibility Study [76].

Antibiotic Resistance and Serotypes/ **Genotypes**

The worldwide increase in antibiotic resistance in S. pneumoniae has been related to the spread of several pneumococcal serotypes (6A, 6B, 9V, 14, 15A, 19F, 19A, and 23F), the socalled 'paediatric serotypes'. In the USA, the CDC's Active Bacterial Core surveillance reported that 24% of 3475 invasive pneumococcal isolates collected in 1998 were PNSP. Seven serotypes (6A, 6B, 9V, 14, 19A, 19F, and 23F) accounted for 91% of all PNSP [64]. After the introduction of PCV7 in 2000 in the USA, a dramatic fall in the incidence of invasive pneumococcal disease (IPD) was observed in children \leq 5 years of age and adults \geq 65 years of age, associated with a decrease in IPD caused by PCV7 serotypes [13,14]. However, an increase in IPD caused by non-PCV7 serotype 19A was observed in all age groups: from 2.6% in 1998–1999 to 47.2% in 2006–2007 in children £5 years of age; from 2.9% to 16.6% in adults 18–64 years of age; and from 3.7% to 14.9% in adults ≥ 65 years of age [14].

In Spain, an increase in the prevalence of combined penicillin and erythromycin resistance among invasive pneumococci was associated with antibiotic consumption in the 1980s and 1990s [16]. Data from the Spanish Reference Laboratory for Pneumococci demonstrated that this increase was related to the rise in prevalence of PCV7 serotypes 6B, 9V, 14, 19F, and 23F, accounting for 76.6% of PNSP in 1979– 1985 and 88% of PNSP in 1998–2000 [16]. The high prevalence of penicillin resistance (36.1%) observed among 5697 invasive pneumococcal isolates collected in Spain in the pre-PCV7 period (1997–2001) was reversed to 22.4% in 2007– 2008 ($n = 5465$) ($p \le 0.001$), when the PCV7 dose distribution increased. On comparison of these two periods, the proportions of four serotypes significantly decreased (p <0.001) among penicillin-resistant strains—serotypes 6B (16.7% vs. 0%), 9V (15.4% vs. 9.1%), 19F (13.7% vs. 7.3%), and 23F (12.6% vs. 3.8%)—whereas the proportion of serotype 14 was similar in both periods (29.6–26.4%). The proportion of non-PCV7 serotypes among PNSP increased from 12.0% in 1997–2001 to 49.5% in 2007–2008, owing to, in particular, significant increases in the proportions of serotypes 19A (3.3–24.5%) and 24F (0.1–7.6%) [16].

The drug-resistant serotypes belong to a small number of pneumococcal clones whose nomenclature is standardized by the Pneumococcal Molecular Epidemiology Network (http:// www.sph.emory.edu/PMEN/) [77]. The most important pneumococcal clones involved in the global spread of antibiotic resistance in the 1980s and 1990s were Spain^{23F}-ST81,

Spain^{6B}-ST90, Spain^{9V}-ST156, England¹⁴-ST9, Taiwan^{19F}-ST236, Taiwan^{23F}-ST242, Poland^{6B}-ST315, Sweden^{15A}-ST63, and Colombia^{23F}-ST338. Although Spain^{23F}-ST81 and Spain^{6B}-ST90 were well-established clones in the 1980s and 1990s, their prevalence decreased after the introduction of PCV7 [78,79]. Spain^{9V}-ST156 (serotypes 9V and 14) has been an important cause of IPD in many countries, before and after PCV7 introduction, and usually shows resistance to penicillin and co-trimoxazole [15,78,79]. The prevalence of England¹⁴-ST9, associated with the spread of macrolide resistance (mefA) in the 1990s in the USA, Canada, the UK, Germany, Norway, Greece, and Italy, decreased significantly after the introduction of PCV7 [44,48,80,81]. The high rate of penicillin and macrolide resistance detected in Asia and the USA was partially related to the spread of Taiwan^{19F}-ST236 [55,82]. The increasing prevalence of the multidrug-resistant serotype 19A observed in the USA is due to the spread of CC320, a double-locus variant of Taiwan^{19F}-ST236 [17]. This serotype 19A clone has also been reported in Asia and Europe [18,19]. However, in Europe, the major clone of serotype 19A is ST276, related to Denmark¹⁴-ST230 [18,20– 22,78].

Antimicrobial use and Antimicrobial **Resistance**

The occurrence of drug-resistant pneumococci has been associated with a variety of factors, antibiotic consumption being one of the most important [25]. Resistance selection has mainly occurred in pneumococci colonizing or infecting children. The frequency of children as carriers, and their exposure to antibiotics, favours the selection of drug-resistant strains [3,4].

Several studies have shown geographical differences in the prevalence of antimicrobial resistance in Europe, with lower rates in the northern countries than in the southern countries [41]. Outpatient antibiotic consumption in the USA was recently compared with data from the European Surveillance Antimicrobial Consumption. Northern European countries reported the lowest antibiotic use, and southern European countries (Greece, France and Italy) the highest, similar to that of the USA. The Netherlands was the country with the lowest reported consumption [12].

Conclusions

S. pneumoniae continues to be an important cause of invasive diseases, especially in children and the elderly. The high

prevalence of multiresistant pneumococci in the 1980s and 1990s declined after the introduction of the PCV7 in the 2000s. However, the emergence of drug-resistant non-PCV7 serotypes/genotypes in the late 2000s emphasizes the importance of prudent use of antibiotics with the aim of preventing their spread. Continued surveillance of antimicrobial resistance, serotypes and genotypes is crucial in providing information on the emergence of multiresistant clones. These data are also essential for the development of appropriate guidelines for empirical therapy of pneumococcal infections and for the inclusion of emergent serotypes in the new generation of conjugate vaccines.

Transparency Declaration

A. Fenoll has received recent research funding from Wyeth. J. Liñares, C. Ardanuy and R. Pallares report no conflicts of interest.

References

- 1. Hansman D, Bullen MM. A resistant pneumococcus. Lancet 1967; 277: 264–265.
- 2. Hansman D, Devitt L, Miles H, Riley I. Pneumococci relatively insensitive to penicillin in Australia and New Guinea. Med J Aust 1974; ii: 353–356.
- 3. Klugman KP. Pneumococcal resistance to antibiotics. Clin Microbiol Rev 1990; 3: 171–196.
- 4. Jacobs MR, Koornhof HJ, Robins-Browne RM et al. Emergence of multiply resistant pneumococci. N Engl J Med 1978; 299: 735–740.
- 5. Liñares J, Garau J, Domínguez C, Pérez JL. Antibiotic resistance and serotypes of Streptococcus pneumoniae from patients with communityacquired pneumococcal disease. Antimicrob Agents Chemother 1983; 23: 545–547.
- 6. Appelbaum PC. World-wide development of antibiotic resistance in pneumococci. Eur J Clin Microbiol 1987; 6: 367–377.
- 7. Michel J, Dickman D, Greenberg Z, Bergner-Rabinowitz S. Serotype distribution of penicillin-resistant pneumococci and their susceptibilities to seven antimicrobial agents. Antimicrob Agents Chemother 1983; 23: 397–401.
- 8. Fenoll A, Martin Bourgon C, Munoz R et al. Serotype distribution and antimicrobial resistance of Streptococcus pneumoniae isolates causing systemic infections in Spain, 1979–1989. Rev Infect Dis 1991; 13: 56– 60.
- 9. Liñares J, Pallares R, Alonso T et al. Trends in antimicrobial resistance of clinical isolates of Streptococcus pneumoniae in Bellvitge Hospital, Barcelona, Spain (1979–1990). Clin Infect Dis 1992; 15: 99– 105.
- 10. Appelbaum PC. Antimicrobial resistance in Streptococcus pneumoniae: an overview. Clin Infect Dis 1992; 15: 77–83.
- 11. Low DE. Changing trends in antimicrobial-resistant pneumococci: it's not all bad news. Clin Infect Dis 2005; 41 (suppl 4): S228–S233.
- 12. Goossens H, Ferech M, Coenen S, Stephens P. European Surveillance of Antimicrobial Consumption Project Group. Comparison of outpatient systemic antibacterial use in 2004 in the United States and 27 European countries. Clin Infect Dis 2007; 44: 1091–1095.
- 13. Kyaw MH, Lynfield R, Schaffner W et al. Effect of introduction of the pneumococcal conjugate vaccine on drug-resistant Streptococcus pneumoniae. N Engl J Med 2006; 354: 1455–1463.
- 14. Pilishvili T, Lexau C, Farley MM et al. Sustained reductions in invasive pneumococcal disease in the era of conjugate vaccine. J Infect Dis 2010; 201: 32–41.
- 15. Aguiar SI, Serrano I, Pinto FR, Melo-Cristino J, Ramirez M. Portuguese Surveillance Group for the Study of Respiratory Pathogens. Changes in Streptococcus pneumoniae serotypes causing invasive disease with non-universal vaccination coverage of the seven-valent conjugate vaccine. Clin Microbiol Infect 2008; 14: 835–843.
- 16. Fenoll A, Granizo JJ, Aguilar L et al. Temporal trends of invasive Streptococcus pneumoniae serotypes and antimicrobial resistance patterns in Spain from 1979 to 2007. J Clin Microbiol 2009; 47: 1012-1020.
- 17. Moore MR, Gertz RE Jr, Woodbury RL et al. Population snapshot of emergent Streptococcus pneumoniae serotype 19A in the United States, 2005. J Infect Dis 2008; 197: 1016–1027.
- 18. Ardanuy C, Rolo D, Fenoll A, Tarrago D, Calatayud L, Liñares J. Emergence of a multidrug-resistant clone (ST320) among invasive serotype 19A pneumococci in Spain. *| Antimicrob Chemother 2009*; 64: 507–510.
- 19. Choi EH, Kim SH, Eun BW et al. Streptococcus pneumoniae serotype 19A in children, South Korea. Emerg Infect Dis 2008; 14: 275–281.
- 20. Dagan R, Givon-Lavi N, Leibovitz E, Greenberg D, Porat N. Introduction and proliferation of multidrug-resistant Streptococcus pneumoniae serotype 19A clones that cause acute otitis media in an unvaccinated population. J Infect Dis 2009; 199: 776–785.
- 21. Mahjoub-Messai F, Doit C, Koeck JL et al. Population snapshot of Streptococcus pneumoniae serotype 19A isolates before and after introduction of seven-valent pneumococcal vaccination for French children. J Clin Microbiol 2009; 47: 837–840.
- 22. Aguiar SI, Pinto FR, Nunes S et al. Increase of Denmark¹⁴-230 clone as a cause of pneumococcal infection in Portugal within a background of diverse serotype 19A lineages. J Clin Microbiol 2010; 48: 101–108.
- 23. Hackenbeck R, Ellerbrok H, Briese T et al. Penicillin-binding proteins of penicillin-susceptible and -resistant pneumococci: immunological relatedness of altered proteins and changes in peptides carrying the β -lactam binding site. Antimicrob Agents Chemother 1986; 30: 553-558.
- 24. Liñares J, Alonso T, Pérez JL et al. Decreased susceptibility of penicillin-resistant pneumococci to twenty-four beta-lactam antibiotics. J Antimicrob Chemother 1992; 30: 279–288.
- 25. Pallares R, Gudiol F, Liñares | et al. Risk factors and response to antibiotic therapy in adults with bacteremic pneumonia caused by penicillin-resistant pneumococci. N Engl J Med 1987; 317: 18–22.
- 26. Pallares R, Linares J, Vadillo M et al. Resistance to penicillin and cephalosporin and mortality from severe pneumococcal pneumonia in Barcelona, Spain. N Engl J Med 1995; 333: 474–480.
- 27. Pallares R, Capdevila O, Liñares | et al. The effect of cephalosporin resistance on mortality in adult patients with nonmeningeal systemic pneumococcal infections. Am J Med 2002; 113: 120–126.
- 28. Friedland IR, Klugman KP. Antibiotic-resistant pneumococcal disease in South African children. Am J Dis Child 1992; 146: 920–923.
- 29. Yu VL, Chiou CC, Feldman C et al. An international prospective study of pneumococcal bacteremia: correlation with in vitro resistance, antibiotics administered, and clinical outcome. Clin Infect Dis 2003; 37: 230–237.
- 30. Song JH, Jung SI, Ko KS et al. High prevalence of antimicrobial resistance among clinical Streptococcus pneumoniae isolates in Asia (an AN-SORP study). Antimicrob Agents Chemother 2004; 48: 2101–2107.
- 31. Weinstein MP, Klugman KP, Jones RN. Rationale for revised penicillin susceptibility breakpoints versus Streptococcus pneumoniae: coping with antimicrobial susceptibility in an era of resistance. Clin Infect Dis 2009; 48: 1596–1600.
- 32. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing: Eighteenth Informational Supplement. M100-S18. Wayne, PA: CLSI, 2008.
- 33. Spika JS, Facklam RR, Plikaytis BD et al. Antimicrobial resistance of Streptococcus pneumoniae in the United States, 1979–1987. The Pneumococcal Surveillance Working Group. J Infect Dis 1991; 163: 1273– 1278.
- 34. Butler JC, Hofmann J, Cetron MS, Elliott JA, Facklam RR, Breiman RF. The continued emergence of drug-resistant Streptococcus pneumoniae in the United States: an update from the Centers for Disease Control and Prevention's Pneumococcal Sentinel Surveillance System. J Infect Dis 1996; 174: 986–993.
- 35. Centers for Disease Control and Prevention.Geographic variation in penicillin resistance in Streptococcus pneumoniae—selected sites, United States, 1997. MMWR 1999; 48: 656–661.
- 36. Felmingham D, White AR, Jacobs MR et al. The Alexander Project: the benefits from a decade of surveillance. J Antimicrob Chemother 2005; 56 (suppl 2): ii3–ii21.
- 37. Jenkins SG, Brown SD, Farrell DJ. Trends in antibacterial resistance among Streptococcus pneumoniae isolated in the USA: update from PROTEKT US Years 1–4. Ann Clin Microbiol Antimicrob 2008; 7: 1–11.
- 38. Felmingham D, Cantón R, Jenkins SG. Regional trends in beta-lactam, macrolide, fluoroquinolone and telithromycin resistance among Streptococcus pneumoniae isolates 2001–2004. J Infect 2007; 55: 111–118.
- 39. Castanheira M, Gales AC, Mendes RE, Jones RN, Sader HS. Antimicrobial susceptibility of Streptococcus pneumoniae in Latin America: results from five years of the SENTRY Antimicrobial Surveillance Program. Clin Microbiol Infect 2004; 10: 645–651.
- 40. Borg MA, Tiemersma E, Scicluna E et al. ARMed Project members and collaborators. Prevalence of penicillin and erythromycin resistance among invasive Streptococcus pneumoniae isolates reported by laboratories in the southern and eastern Mediterranean region. Clin Microbiol Infect 2009; 15: 232–237.
- 41. European Antimicrobial Resistance Surveillance System. EARSS Annual Report 2008. Available at: http://www.rivm.nl/earss/Images/EARSS% 202008 final tcm61-65020.pdf (last accessed 26 November 2009).
- 42. Doern GV, Heilmann KP, Huynh HK et al. Antimicrobial resistance among clinical isolates of Streptococcus pneumoniae in the United States during 1999–2000, including a comparison of resistance rates since 1994–1995. Antimicrob Agents Chemother 2001; 45: 1721–1729.
- 43. Richter SS, Heilmann KP, Dohrn CL, Riahi F, Beekmann SE, Doern GV. Changing epidemiology of antimicrobial-resistant Streptococcus pneumoniae in the United States, 2004–2005. Clin Infect Dis 2009; 48: e23–e33.
- 44. van der Linden M, Al-Lahham A, Haupts S, Reinert RR. Clonal spread of mef-positive macrolide-resistant Streptococcus pneumoniae isolates causing invasive disease in adults in Germany. Antimicrob Agents Chemother 2007; 51: 1830–1834.
- 45. Amezaga MR, Carter PE, Cash P, McKenzie H. Molecular epidemiology of erythromycin resistance in Streptococcus pneumoniae isolates from blood and noninvasive sites. J Clin Microbiol 2002; 40: 3313–3318.
- 46. Littauer P, Sangvik M, Caugant DA, Høiby EA, Simonsen GS, Sundsfjord A. Norwegian Macrolide Study Group. Molecular epidemiology of macrolide-resistant isolates of Streptococcus pneumoniae collected from blood and respiratory specimens in Norway. | Clin Microbiol 2005; 43: 2125–2132.
- 47. Van Eldere J, Meekers E, Lagrou K et al. Macrolide-resistance mechanisms in Streptococcus pneumoniae isolates from Belgium. Clin Microbiol Infect 2005; 11: 332–334.
- 48. Monaco M, Camilli R, D'Ambrosio F, Del Grosso M, Pantosti A. Evolution of erythromycin resistance in Streptococcus pneumoniae in Italy. J Antimicrob Chemother 2005; 55: 256–259.
- 49. Reinert RR, Ringelstein A, van der Linden M, Cil MY, Al-Lahham A, Schmitz FJ. Molecular epidemiology of macrolide-resistant Streptococcus

pneumoniae isolates in Europe. J Clin Microbiol 2005; 43: 1294– 1300.

- 50. Siira L, Rantala M, Jalava J et al. Temporal trends of antimicrobial resistance and clonality of invasive Streptococcus pneumoniae isolates in Finland, 2002 to 2006. Antimicrob Agents Chemother 2009; 53: 2066–2073.
- 51. Calatayud L, Ardanuy C, Cercenado E et al. Serotypes, clones, and mechanisms of resistance of erythromycin-resistant Streptococcus pneumoniae isolates collected in Spain. Antimicrob Agents Chemother 2007; 51: 3240–3246.
- 52. Reinert RR, Filimonova OY, Al-Lahham A et al. Mechanisms of macrolide resistance among Streptococcus pneumoniae isolates from Russia. Antimicrob Agents Chemother 2008; 52: 2260–2262.
- 53. Bae S, Lee K. Distribution of capsular serotypes and macrolide resistance mechanisms among macrolide-resistant Streptococcus pneumoniae isolates in Korea. Diagn Microbiol Infect Dis 2009; 63: 213–216.
- 54. Isozumi R, Ito Y, Ishida T et al. Genotypes and related factors reflecting macrolide resistance in pneumococcal pneumonia infections in Japan. J Clin Microbiol 2007; 45: 1440–1446.
- 55. Song JH, Chang HH, Suh JY et al. Macrolide resistance and genotypic characterization of Streptococcus pneumoniae in Asian countries: a study of the Asian Network for Surveillance of Resistant Pathogens (ANSORP). J Antimicrob Chemother 2004; 53: 457–463.
- 56. Jenkins SG, Farrell DJ. Increase in pneumococcus macrolide resistance, United States. Emerg Infect Dis 2009; 15: 1260–1264.
- 57. Wierzbowski AK, Nichol K, Laing N et al. Macrolide resistance mechanisms among Streptococcus pneumoniae isolated over 6 years of Canadian Respiratory Organism Susceptibility Study (CROSS) (1998– 2004). J Antimicrob Chemother 2007; 60: 733–740.
- 58. Wolter N, von Gottberg A, du Plessis M, de Gouveia L, Klugman KP. Group for Enteric, Respiratory Meningeal Disease Surveillance in South Africa Molecular basis and clonal nature of increasing pneumococcal macrolide resistance in South Africa, 2000–2005. Int J Antimicrob Agents 2008; 32: 62–67.
- 59. Farrell DJ, Jenkins SG, Brown SD, Patel M, Lavin BS, Klugman KP. Emergence and spread of Streptococcus pneumoniae with erm(B) and mef(A) resistance. Emerg Infect Dis 2005; 11: 851–858.
- 60. Del Grosso M, Northwood JG, Farrell DJ, Pantosti A. The macrolide resistance genes erm(B) and mef(E) are carried by Tn2010 in dualgene Streptococcus pneumoniae isolates belonging to clonal complex CC271. Antimicrob Agents Chemother 2007; 51: 4184–4186.
- 61. Buu-Hoï AY, Goldstein FW, Acar JF. A seventeen-year epidemiological survey of antimicrobial resistance in pneumococci in two hospitals. J Antimicrob Chemother 1988; 22 (suppl B): 41–52.
- 62. Doern GV, Brown SD. Antimicrobial susceptibility among community acquired respiratory tract pathogens in the USA: data from PRO-TEKT US 2000–01. J Infect 2004; 48: 56–65.
- 63. Pérez-Trallero E, Fernández-Mazarrasa C, García-Rey C et al. Antimicrobial susceptibilities of 1,684 Streptococcus pneumoniae and 2,039 Streptococcus pyogenes isolates and their ecological relationships: results of a 1-year (1998–1999) multicenter surveillance study in Spain. Antimicrob Agents Chemother 2001; 45: 3334–3340.
- 64. Whitney CG, Farley MM, Hadler J et al. Increasing prevalence of multidrug-resistant Streptococcus pneumoniae in the United States. N Engl J Med 2000; 343: 1917–1924.
- 65. Lonks JR, Garau J, Gomez L et al. Failure of macrolide antibiotic treatment in patients with bacteremia due to erythromycin-resistant Streptococcus pneumoniae. Clin Infect Dis 2002; 35: 556–564.
- 66. Marimón JM, Iglesias L, Vicente D, Pérez-Trallero E. Molecular characterization of erythromycin-resistant clinical isolates of the four major antimicrobial-resistant Spanish clones of Streptococcus pneumo-

niae (Spain^{23F}-1, Spain^{6B}-2, Spain^{9V}-3, and Spain¹⁴-5). Microb Drug Resist 2003; 9: 133–137.

- 67. Muñoz R, De La Campa AG. ParC subunit of DNA topoisomerase IV of Streptococcus pneumoniae is a primary target of fluoroquinolones and cooperates with DNA gyrase A subunit in forming resistance phenotype. Antimicrob Agents Chemother 1996; 40: 2252– 2257.
- 68. de la Campa AG, Balsalobre L, Ardanuy C, Fenoll A, Pérez-Trallero E, Liñares J. Spanish Pneumococcal Infection Study Network G03/103. Fluoroquinolone resistance in penicillin-resistant Streptococcus pneumoniae clones, Spain. Emerg Infect Dis 2004; 10: 1751– 1759.
- 69. de la Campa AG, Ardanuy C, Balsalobre L et al. Changes in fluoroquinolone-resistant Streptococcus pneumoniae after 7-valent conjugate vaccination, Spain. Emerg Infect Dis 2009; 15: 905–911.
- 70. Chen DK, McGeer A, De Azavedo JC, Low DE. Decreased susceptibility of Streptococcus pneumoniae to fluoroquinolones in Canada. N Engl J Med 1999; 341: 233–239.
- 71. Liñares I, de la Campa AG, Pallares R. Fluoroguinolone resistance in Streptococcus pneumoniae. N Engl J Med 1999; 341: 1546–1547.
- 72. Riedel S, Beekmann SE, Heilmann KP et al. Antimicrobial use in Europe and antimicrobial resistance in Streptococcus pneumoniae. Eur J Clin Microbiol Infect Dis 2007; 26: 485–490.
- 73. Fuller JD, Low DE. A review of Streptococcus pneumoniae infection treatment failures associated with fluoroquinolone resistance. Clin Infect Dis 2005; 41: 118–121.
- 74. Adam HJ, Hoban DJ, Gin AS, Zhanel GG. Association between fluoroquinolone usage and a dramatic rise in ciprofloxacin-resistant Streptococcus pneumoniae in Canada, 1997–2006. Int J Antimicrob Agents 2009; 34: 82–85.
- 75. Carratalá J, Martín-Herrero JE, Mykietiuk A, García-Rey C. Clinical experience in the management of community-acquired pneumonia: lessons from the use of fluoroquinolones. Clin Microbiol Infect. 2006; 12: (suppl 3):2–11.
- 76. Schurek KN, Adam HJ, Siemens CG, Hoban CJ, Hoban DJ, Zhanel GG. Are fluoroquinolone-susceptible isolates of Streptococcus pneumoniae really susceptible? A comparison of resistance mechanisms in Canadian isolates from 1997 and 2003. J Antimicrob Chemother 2005; 56: 769–772.
- 77. McGee L, McDougal L, Zhou J et al. Nomenclature of major antimicrobial-resistant clones of Streptococcus pneumoniae defined by the pneumococcal molecular epidemiology network. J Clin Microbiol 2001; 39: 2565–2571.
- 78. Ardanuy C, Tubau F, Pallares R et al. Epidemiology of invasive pneumococcal disease among adult patients in Barcelona before and after pediatric 7-valent pneumococcal conjugate vaccine introduction, 1997–2007. Clin Infect Dis 2009; 48: 57–64.
- 79. Zhou J, Enright MC, Spratt BG. Identification of the major Spanish clones of penicillin-resistant pneumococci via the Internet using multilocus sequence typing. J Clin Microbiol 2000; 38: 977–986.
- 80. Fotopoulou N, Tassios PT, Beste DV et al. A common clone of erythromycin-resistant Streptococcus pneumoniae in Greece and the UK. Clin Microbiol Infect 2003; 9: 924–929.
- 81. McEllistrem MC, Adams JM, Shutt K et al. Erythromycin-nonsusceptible Streptococcus pneumoniae in children, 1999–2001. Emerg Infect Dis 2005; 11: 969–972.
- 82. Gertz RE Jr, McEllistrem MC, Boxrud DJ et al. Clonal distribution of invasive pneumococcal isolates from children and selected adults in the United States prior to 7-valent conjugate vaccine introduction. J Clin Microbiol 2003; 41: 4194–4216.